

Total Phenol Content, Total Flavonoid Content, Antioxidant Activity and GC-MS Analysis of a New Variety of Rice (*Oryza sativa* L.)

Hafidha Asni Akmalia*, Dwimei Ayudewandari Pranatami

Biology Education Department, Faculty of Science and Technology, Universitas Islam Negeri Walisongo Semarang,
Jl. Prof. Dr. Hamka, Ngaliyan, Semarang 50185, Tel. (024) 7604554, Indonesia.

Corresponding author*

akmalia.h.a@walisongo.ac.id

Manuscript received: 16 November, 2025. Revision accepted: 18 December, 2025. Published: 02 January, 2026.

Abstract

Consuming foods rich in antioxidants plays an important role in human health, one of which is through rice. This research aimed to describe the total phenol content and total flavonoids content as well as GC-MS analysis of raw Baroma rice extract and analyze the antioxidant activity of Baroma rice in different cooking methods. Baroma rice was collected from Semarang, Central Java, Indonesia thus was analyzed to get data about total phenol content, total flavonoid content, and compounds identifies using GC-MS. We also analyzed antioxidant activity of Baroma rice among raw, rice cooked in stove and *magic com* extract using Anova test followed by Tukey test. The result proved that total phenol content was 85.64 mg/L and total flavonoid content was 78.28 mg/L, meanwhile the highest antioxidant activity was in raw extract of Baroma followed by rice cooked in magic com and stove, suggesting that cooking method effected on its property. GC-MS analysis showed the diversity of chemical compound, totaling 15 compounds and peaks.

Keywords: antioxidant activity; Baroma variety; GC-MS analysis; flavonoid; phenol.

INTRODUCTION

Food is an essential nutrient for humans, especially its nutritional content for growth. Food is also actually part of the treatment that can be done for health. Over time, the existence of EPFRs which is abbreviation of environmentally persistent free radicals has become a health threat due to their potential to damage DNA and be toxic (Qin et al., 2025). Therefore, consuming foods containing antioxidants is one way to neutralize the malignant effects of free radicals. This requires vigilance, considering that free radicals such as ROS (hydrogen peroxide (H₂O₂), hydroxyl radicals (•OH), (superoxide radicals (O₂•-), and singlet oxygen (1O₂)) are suspected of causing diseases (Pizzino et al., 2017) such as cancer (R. Huang et al., 2021), diabetes (Volpe et al., 2018), and cardiovascular disease (Elias-Llumbet et al., 2024).

Antioxidant activity has been studied in many plant species such as the Apiaceae family (Bello et al., 2025), *Bauhinia saccocalyx* (Katisart et al., 2025), *Bacopa monnieri* (Govindarasu & Murugesh, 2025), *Schefflera vinosa* (Singh et al., 2021). In *Oryza sativa*, its antioxidant activity is also worth considering that rice is a staple food consumed by more than half of the world's population, especially in Asia mainly due to its protein, carbohydrate, vitamin, and essential mineral content (Colasanto et al., 2024). In colored rice varieties such as black rice western Taiwan; Taibalang red waxy rice;

Taibalang black waxy rice; black rice Thailand; red rice Thailand; Guangfu red rice; are proven to have high antioxidant activity (Y. P. Huang & Lai, 2016). In line with Huang, (Shao et al., 2014) also stated that pigmented rice, especially black rice, has higher antioxidant activity and total phenolic content in the endosperm, embryo, and bran compared to white rice and red rice. Brown and purple rice are also reported to have promising antioxidants as in the study (Gong et al., 2017) (Li et al., 2024). This antioxidant activity is closely related to the existence of phenols and flavonoids (Suryanti et al., 2020). (Yusoff et al., 2022) argued that flavonoids are bioactive compounds that contribute to antioxidant activity.

Baroma is a rice variety launched directly by the Indonesian Ministry of Agriculture in 2019 through Balai Besar Padi (Rice Center in Indonesia). Baroma is an aromatic Basmati rice that has been refined by Indonesian researchers consequently it can grow in lowlands. Baroma is claimed to have higher productivity compared to Basmati and the rice size remains long even when planted in lowlands. Research on Baroma rice is still limited, such as (Pranatami & Akmalia, 2025)'s research on the potential of Baroma rice as an anticancer, the nutritional value of Baroma rice in different cooking methods (Akmalia et al., 2024), and Baroma rice as a variety for food security (Windiyani & Rusdianto, 2020).

Information related to metabolite or compounds and the total content of Baroma rice has never been reported. Therefore, an in-depth analysis of total phenols, total flavonoids and GC-MS analysis of raw Baroma rice as well as antioxidant activity with different cooking methods is necessary. This study aims to describe the total phenol content and total flavonoids content as well as GC-MS analysis of raw Baroma rice extract and analyze the antioxidant activity of Baroma rice in different cooking methods.

MATERIALS AND METHODS

Materials

The materials used in this research were Baroma rice (*Oryza sativa* L.) collected from Semarang, Central Java, Follin ciocalteus phenol reagent, aquadest, gallic acid, sodium nitrite, aluminum nitrate, sodium hydroxide, quercetin, methanol (Merck), DPPH reagent, ethanol (Merck). The tools used were food thermometer, *magic com*, stove, spectrophotometer, and Thermo Scientific GC-MS.

Procedures

Preparation of tested rice

Two hundred g of rice (*Oryza sativa* L.) was cooked using a stove and another 200 g was cooked using a *magic com* in 20 minutes. The temperature of the cooked rice was measured using a food thermometer with the results of the stove-cooked rice having a temperature of 87.8°C, and 87.9°C for the *magic com*-cooked rice. We tested the total phenols and flavonoids in the raw rice extract and for the further test, we did the antioxidant test. Baroma rice extract was obtained through the maceration method with 100 mL of ethanol solvent using 200 g of raw Baroma rice, rice cooked in *magic com*, and rice cooked in stove. Antioxidant activity testing was carried out on 3 extracts of raw Baroma rice, cooked *magic com*, and cooked stove using the DPPH method. We also tested the compound in raw extract using Gas Chromatography-Mass spectrometry (GC-MS).

Determination of Total Phenol Content Equivalent Gallic Acid using Spectrophotometric Method

A sample of 1 mL was taken then add 0.5 mL of Follin ciocalteus phenol reagent and 7.5 mL of Aquadest. It was shaken then let it stand for 10 minutes. We added 1.5 mL of 20% sodium carbonate solution and shake also let it stand for 10 minutes. Add Aquadest up to 10 mL and read the absorbance at a wavelength of 760 nm.

Determination of Total Flavonoid Content Equivalent Quercetin using Spectrophotometric Method

A sample of 1 mL was taken then add 0.3 mL of 5% sodium nitrite and wait for 5 minutes. Add 0.6 mL of 10% aluminum nitrate, wait for 5 minutes. It was then added 2 mL of 1 M sodium hydroxide and adjust it to a

volume of 10 mL. The last step was the reading of absorbance at λ 510 nm.

Antioxidant Analysis Procedure by DPPH Method Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995) Method (Brand-Williams et al., 1995)

Five mL PA methanol was added to sample 0.2 g then extracted by vortexing. The extract was taken as much as 0.2 mL. Add 2.8 mL of 0.1 mM DPPH reagent, then incubated at room temperature and dark room for 1 hour. It was then calibrated with a 515 nm spectrophotometer, also did it to make a blank of 0.2 mL methanol + 2.8 mL of 0.1 mM DPPH reagent.

The DPPH free radical scavenging activity was calculated as follows and was measured equivalent in % wb:

$$\% \text{ DPPH} = \frac{1 - \text{sample absorbance}}{\text{sample absorbance}} \times 100\%$$

GC-MS Analysis Procedure

The ethanol extract of raw Baroma rice was analyzed by Thermo Scientific GC-MS. It was run using column with length 30 m, diameter 0,250 mm, and film thickness 0,25 μm . Injector temperature was 250°C, splitless injection method, and the carrier gas was helium with operating pressure of 70,69 kPa. An initial temperature we used was 40°C for 3 min; a linear increase to 100°C at 5°C/minutes and hold for 3 min and ramping to 250°C at 10°C/ minutes and hold for 4 minutes (Hu et al., 2023).

Data analysis

Data related to total phenol content, total flavonoid content, and GC-MS analysis were described descriptively while the antioxidant activity was measured equivalent in % wb then the values obtained from the three extracts were analyzed using the ANOVA test using JASP software followed by Tukey test.

RESULTS AND DISCUSSION

We tested the presence of flavonoid and phenolic compounds, which play a role in scavenging free radicals, by testing total phenols and flavonoids. Groups of bioactive substances known as flavanoids are further classified as flavones, flavanols, flavanones, flavonols, and isoflavones. The results showed that the total observed phenol was 85.64 mg/L and the total flavonoid was 78.28 mg/L (Table 1).

Table 1. Total phenol dan flavonoid content in raw Baroma rice.

No	Parameter	Calculated Amount (mg/L)
1.	Total phenol content	85,64
2.	Total flavonoid content	78,28

The results of the antioxidant activity of the raw extract and the *magic com* and stove methods are described in Table 2. Cooking significantly reduces antioxidant activity, with the lowest value in Baroma rice cooked using a stove. However, % wb values for both cooking methods are still relatively high, so these methods can be recommended for use when cooking rice.

Table 2. Antioxidant Activity of Baroma rice in raw, magic com cooking and stove cooking.

No	Extract	% wb
1.	Raw Baroma	2,23 ^a
2.	Magic com cooking of Baroma	1,74 ^{ab}
3.	Stove cooking of Baroma	1,08 ^b

Numbers followed by the same letter indicate no significant difference in the Tukey test with a 95% confidence level.

GC-MS analysis is performed to detect compounds even in small concentrations. The advantages of GC-MS include its high sensitivity and the ability to separate mixed compounds (Arsana et al., 2022), although it depends on extraction conditions, sample preparation techniques, and analytical parameters (Phuong et al., 2025). The result of GC-MS detection is summarized in Table 3.

Table 3. Compound identified in Baroma Rice using GC-MS.

Peak	Retention Time	Similarity Index	Compound Name	Relative Area (%)	Formula
7	4,93	731	Ethanone, 2-(formyloxy)-1-phenyl-	5,87	C9H8O3
11	6,58	819	Arsenous acid, tris(trimethylsilyl) ester	0,62	C9H27AsO3Si3
20	24,46	867	1-Dodecanol	0,24	C12H26O
21	24,52	928	Cyclododecane	3,23	C12H24
23	25,11	911	2,4-Di-tert-butylphenol	0,70	C14H22O
24	26,19	869	Dodecanoic acid, ethyl ester	0,38	C14H28O2
27	27,40	819	2-Propenoic acid, pentadecyl ester	6,08	C18H34O2
36	28,53	877	Tetradecanoic acid, ethyl ester	1,04	C16H32O2
38	29,45	788	5-Octadecenal	0,17	C18H34O
40	30,58	895	Hexadecanoic acid, ethyl ester	16,93	C18H36O2
43	30,87	809	Isopropyl palmitate	0,27	C19H38O2
44	32,17	900	n-Propyl 9,12-octadecadienoate	8,32	C21H38O2
45	32,21	871	Ethyl Oleate	6,06	C20H38O2
46	32,27	808	n-Propyl 11-octadecenoate	0,94	C21H40O2
47	32,43	836	Eicosanoic acid, ethyl ester	8,57	C22H44O2

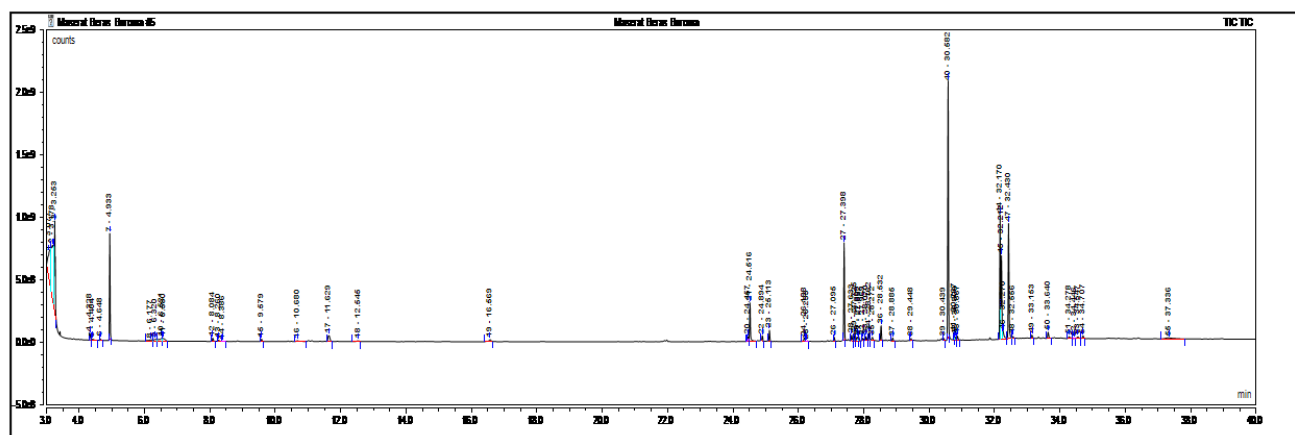


Figure 1. Chromatogram of GC-MS analysis.

Figure 1 shows that there are 55 peaks and 133 compounds of Baroma rice analyzed using GC-MS. After sorting the compounds through Similarity Index selection and comparison with the PubChem database, 15

compounds were successfully identified. These compounds were located at peaks 7, 11, 20, 21, 23, 24, 27, 36, 38, 40, 43, 44, 45, 46, and 47. As clearly seen in

Table 3 and Figure 1, peak number 40 has the largest relative area of 16.93% with the retention time is 30,582.

Discussion

Baroma rice (*Oryza sativa* L.) is white rice which, when compared to pigmented rice, apparently has fewer total phenols and flavonoids (see Table 1). This is proven in the study of (Ghasemzadeh et al., 2018) who obtained a total phenolic content of black rice of 1214.7 mg GAE/100 g DM and 811.32 mg GAE/100 g DM for red rice. Meanwhile, the total flavonoid content of black rice was mg QE/100 g DM and red rice was 457.00 mg QE/100 g DM. Phenols and flavonoids have been known as potential antioxidants in black rice (Suryanti et al., 2020) which play a role in capturing free radicals such as ROS and RNS thus they can be used for antimicrobial, anti-inflammatory, anticarcinogenic and anticancer therapy. Free radicals can damage cell components such as fat, protein, nucleic acid and ultimately cause cell death (Woo et al., 2018). (Subedi et al., 2014) claimed that free radical production induces disorders such as asthma, diabetes, cardiovascular disease, cancer, inflammatory disease, neurodegenerative diseases, and premature aging. Phenols and flavonoids work by directly scavenging ROS radicals and inhibiting ROS formation by chelating trace elements, such as quercetin, which chelates iron and stabilizes iron (Dias et al., 2021). According to (Banjamahor & Artanti, 2014), the free hydroxyl group will provide a radical molecule its hydrogen atom, stabilizing it and producing a relatively stable flavonoid phenoxyl radical. The presence of phenol and flavonoid compounds is very linear with the antioxidant activity of plants (Kusumawardani & Luangsakul, 2024; Subedi et al., 2014).

Das et al.(2023)contended that knowledge about food nutrition is important for regular consumption. The decrease in antioxidant activity in cooked rice (see Table 2) is thought to be due to a decrease in phenol or flavonoid levels. (Fracassetti et al., 2020) studied the antioxidant activity of raw and cooked Carnaroli, Venere, and Artemide rice varieties. The results showed that the Total Phenol Index decreased to 31.7% in Carnaroli, 58.3% in Venere, and 71.7% in Artemide compared to the contents in the raw samples. Similar results were obtained by (Phanthurat & Thatsanasuwan, 2023), where the streamer and rice cooker methods significantly reduced total phenol and flavonoid content, as well as antioxidant activity. The reduction of phenolic compounds following cooking can be ascribed to various factors, including the water-rice ratio, the duration of cooking (Fracassetti et al., 2020), and temperatures (Aleixandre et al., 2022). Furthermore, the loss of total phenol content (TPC) may be due to several factors including the migration of biocompounds into the cooking water, chemical degradation or transformation into different compounds, as well as interactions with

other food components, which consequently diminishing their extractability (Bani et al., 2024).

Bani et al. (2024) also emphasized that cooking can lead to a loss of antioxidant activity, observed in rice varieties BO (brown orange); BC (brown carnaroli); BV (brown violet); MC (milled carnaroli); MV (milled violet); MO (milled orange); The varieties with the most significant reduction in AOA were Brown violet and milled carnaroli. Furthermore, (Tang et al., 2016)'s research found that anthocyanins, which also play a role in antioxidant activity, undergo degradation during high-temperature cooking using the rice cooker method. Consuming food with proper cooking methods can prevent the loss of antioxidants. Elevated level of ROS facilitate tumor progression by impairing the function of T cells and natural killer (NK) cells while also encouraging the recruitment and M2 polarization of macrophages (R. Huang et al., 2021). Therefore, it is necessary to adjust rice cooking methods to maintain high antioxidant activity.

As observed in Figure 1 and Table 3, the chromatogram shows that there are 15 peaks volatile compounds successfully extracted from raw Baroma rice. The compound in peak 40 is identified to be Hexadecanoic acid, ethyl ester which has the highest composition characterized by a relative area of 16.93% and a retention time of 30.58. Hexadecanoic acid, ethyl ester compound has a molecular weight of 284.5 g/mol and a molecular formula of C₁₈H₃₆O₂. This compound has been proven to have potential as an anti-inflammatory tested on model mice (Saeed et al., 2012), nematocide or pesticide (Sulochana et al., 2016) and anti-chikungunya virus (Sagna et al., 2023). In this peak, another compound actually is suspected to be Pentadecanoic acid, ethyl ester that has a molecular weight of 270.5 g/mol and a molecular formula of C₁₇H₃₄O₂. This compound is a derivative of palmitic acid, which is a significant element influencing the efficacy of chemotherapy in multiple myeloma (Dong et al., 2024). (Casillas-Vargas et al., 2021) also reported that this compound has antibacterial activity against both gram-positive and gram-negative bacteria. At peak 47, it is proved to be Eicosanoic acid, ethyl ester with a relative area of 8.57% and a retention time of 32.430. Eicosanoic acid, ethyl ester is a compound possessing a molecular weight of 340.6 g/mol and a molecular formula of C₂₂H₄₄O₂ which is formed by the esterification of long-chain straight-chain fatty acids, eicosanoic acid and ethanol. (Sokeng et al., 2020) showed that Eicosanoic acid, ethyl ester acts as an anti-inflammatory and analgesic effect.

n-Propyl 9,12-octadecadienoate is a compound in peak 44. The compound n-Propyl 9,12-octadecadienoate, also known as linoleic acid propyl ester, has a molecular weight of 322.5 g/mol and a molecular formula of C₂₁H₃₈O₂. This compound is an ester of a fatty acid resulting from the formal condensation of the carboxylic

group of inoleic acid with the hydroxy group of propanol. Linoleic acid is the main active ingredient in Wali fruit seed oil which has various biological and pharmacological activities, and is often used to cure various cancers, such as liver, lung, and gastrointestinal cancer (Wu et al., 2017). Besides, another suspected compound in this peak is 9,12-Octadecadienoic acid that has a molecular weight of 280.4 g/mol and a molecular formula of C₁₈H₃₂O₂. 9,12-Octadecadienoic acid is otherwise known as omega-6 unsaturated fatty acid (Krishnaveni et al., 2014). It is also apparently useful in the treatment of hyperlipidemia and atherosclerosis (Hamilton-Amachree & Uzoekwe, 2017) and has proven antibacterial and antifungal activity against *S. aureus*, *S. pyogenes*, *S. typhi*, *E. coli*, *C. albicans*, and *A. niger* (Onoabedje et al., 2025).

Arsenous acid, tris(trimethylsilyl) ester in Baroma rice was observed at peak 11. Recent studies outlined by (Wiraswati et al., 2023) suggest that this compound was also seen in methanolic extract of *Momordica cymbalaria* containing biological activity to combat diabetes mellitus, diarrhea, skin disease, rheumatism, and ulcers. Cyclododecane, as found in peak 21, is a member of cycloalkanes and 2,4-Di-tert-butylphenol in peak 23 is the property of the class of organic compounds known as phenylpropanes. Dodecanoic acid, ethyl ester also known as ethyl laurate or ethyl laurate belongs to fatty acid esters. This compound is a multipurpose emulsifier, moisturizer, and texture-enhancing ingredient in the cosmetics industry (Villanueva-Bermejo et al., 2025). Meanwhile, (Khromykh et al., 2022) found that 2-Propenoic acid, pentadecyl ester express antioxidant and antimicrobial activity in fruits and leaves extract of *Actinidia polygama* and *A. arguta*. Tetradecanoic acid, ethyl ester also known as Ethyl myristate, makes up a significant portion of the total fatty acids in palm kernel and coconut oils (Mohebbifar, 2021). It is a part of the fatty acid ethyl esters and is made of myristic acid. It is also used as one of biodiesels component. As established by (Guo et al., 2006), Isopropyl palmitate acts as a penetration enhancer of drugs in ethanol solution and (Chen et al., 2021) claimed that Ethyl oleate as effective acaricides for the control of spider mites. Moreover, n-Propyl 11-octadecenoate reported in *S. mombin* leaf extract acts as an organic addition in the production of biogenic soap (Ohiri et al., 2023). The research also showed the presence of 5-octadecenal in peak 38 as aldehydes. Aldehydes are odorous compounds commonly found in fragrant rice, such as Baroma rice, and are typically synthesized from the oxidation of free fatty acids or the decomposition of linoleic acid (Hu et al., 2023). Besides, Ethanone, 2-(formyloxy)-1-phenyl- in peak 7 also produces aromatic compound in the presence of ketone. Furthermore, 1-Dodecanol, a fatty alcohol, was detected, which also plays a role in producing floral odors.

The production of metabolite compounds is highly dependent on the region (Arsana et al., 2022), climate

change, and environmental conditions around the plant (Pant et al., 2021). (Feng et al., 2019)'s research related to GC-MS analysis proves that secondary metabolite compounds in rice in different habitats have differences. Rice from the Jiansanjiang region has unique compounds, there are benzoic acid, glucose, fumaric acid, Myo-inositol, Xylose, xylitol, sorbitol, and Raffinose, while rice from Wuchang has Pentanoic acid, Glucopyranose, Octadecanoic acid, Eicosanoic acid, Docosanoic acid, 1-Monohexadecanoylglycerol, Trehalose, β -Tocopherol, and β -sitosterol. In this study, Baroma rice analyzed using GC-MS came from Semarang, Central Java, so it is very possible that the results will be different if using Baroma rice grown in other areas. Research related to Baroma rice can still be elaborated again with the profiling of metabolite compounds and antioxidant activity in rice with different habitat.

CONCLUSIONS

The total phenol and flavonoid content of Baroma rice were detected at 85.64 mg/L and 78.28 mg/L, respectively. The highest antioxidant activity was observed in the raw extract of Baroma rice, while the magic com cooking method had a higher % wb value than the stovetop cooking method. Meanwhile, GC-MS analysis showed that 15 compounds were successfully extracted, with peak number 40 having the largest relative area of 16.93%. The compound is identified to be hexadecanoic acid, ethyl ester. Further research can be conducted, including analyzing other compounds that play a role in antioxidant activity such as carotenoids, vitamins, or trace elements and expanding the variety of cooking methods to find the most appropriate method for maintaining the antioxidant activity of Baroma rice.

Acknowledgements: The authors thank to Ministry of Religious Affairs for the research grants provided and Laboratory of Structure and Development of Universitas Islam Negeri Walisongo Semarang for the facility.

Authors' Contributions: Hafidha Asni Akmalia and Dwime Ayudewandari designed the study. Hafidha Asni Akmalia carried out the laboratory work. Hafidha Asni Akmalia and Dwime Ayudewandari analyzed the data. Hafidha Asni Akmalia wrote the manuscript. All authors read and approved the final version of the manuscript.

Competing Interests: The authors declare that there are no competing interests.

REFERENCES

Akmalia, H. A., Febriana, A., Pranatami, D. A., & Adawiyah, P. R. (2024). Nutritional Status of Baroma Rice in Different

- Cooking Methods. *Jurnal Gizi Dan Pangan Soedirman*, 8(2), 204. <https://doi.org/10.20884/1.jgipas.2024.8.2.12808>
- Aleixandre, A., Gil, J. V., Sineiro, J., & Rosell, C. M. (2022). Understanding phenolic acids inhibition of α -amylase and α -glucosidase and influence of reaction conditions. In *Food Chemistry* (Vol. 372). <https://doi.org/10.1016/j.foodchem.2021.131231>
- Arsana, I. N., Juliasih, N., Ayu Sauca Sunia Widyantari, A., Suriani, N., & Manto, A. (2022). GC-MS Analysis of the Active Compound in Ethanol Extracts of White Pepper (*Piper nigrum* L.) and Pharmacological Effects. *Cellular, Molecular and Biomedical Reports*, 2(3), 151–161. <https://doi.org/10.55705/cnbr.2022.351720.1051>
- Bani, C., Cappa, C., Restani, P., Sala, M., Colombo, F., Mercogliano, F., & Di Lorenzo, C. (2024). Physicochemical and nutritional quality of pigmented rice and bran: Influence of milling and cooking. *Lwt*, 208(April). <https://doi.org/10.1016/j.lwt.2024.116653>
- Banjarnahor, S. D. S., & Artanti, N. (2014). Antioxidant properties of flavonoids. *Medical Journal of Indonesia*, 23(4), 239–244. <https://doi.org/10.13181/mji.v23i4.1015>
- Bello, A. A., Issa, N., Mawardi, K., & Batch, A. (2025). Antioxidant Activity of Some Apiaceae Plants Wild Distributed in Aleppo, Syria. *South African Journal of Chemical Engineering*, 54(August), 200–209. <https://doi.org/10.1016/j.sajce.2025.08.003>
- Brand-Williams, W., Cuvelier, M. E., & Berset, C. (1995). Use of a free radical method to evaluate antioxidant activity. *LWT - Food Science and Technology*, 28(1), 25–30. [https://doi.org/10.1016/S0023-6438\(95\)80008-5](https://doi.org/10.1016/S0023-6438(95)80008-5)
- Casillas-Vargas, G., Ocasio-Malavé, C., Medina, S., Morales-Guzmán, C., Del Valle, R. G., Carballeira, N. M., & Sanabria-Ríos, D. J. (2021). Antibacterial fatty acids: An update of possible mechanisms of action and implications in the development of the next-generation of antibacterial agents. *Progress in Lipid Research*, 82(January). <https://doi.org/10.1016/j.plipres.2021.101093>
- Chen, Y., Luo, J., Wan, N., Jiang, J., & Dai, G. (2021). Carmine spider mite *Tetranychus cinnabarinus* control: Laboratory and field efficacy and biochemical characterization of 2, 4-di-tertbutylphenol and ethyl oleate. *Crop Protection*, 139(September 2020), 105390. <https://doi.org/10.1016/j.cropro.2020.105390>
- Colasanto, A., Travaglia, F., Bordiga, M., Coïsson, J. D., Arlorio, M., & Locatelli, M. (2024). Impact of traditional and innovative cooking techniques on Italian black rice (*Oryza sativa* L., Artemide cv) composition. *Food Research International*, 194(June). <https://doi.org/10.1016/j.foodres.2024.114906>
- Das, M., Dash, U., Mahanand, S. S., Nayak, P. K., & Kesavan, R. K. (2023). Black rice: A comprehensive review on its bioactive compounds, potential health benefits and food applications. *Food Chemistry Advances*, 3(September). <https://doi.org/10.1016/j.focha.2023.100462>
- Dias, M. C., Pinto, D. C. G. A., & Silva, A. M. S. (2021). Plant flavonoids: Chemical characteristics and biological activity. *Molecules*, 26(17), 1–16. <https://doi.org/10.3390/molecules26175377>
- Dong, H., Chen, J., Zhang, H., Zhao, M., Yue, Y., & Wang, S. (2024). Palmitic acid inhibits macrophage-mediated chemotherapy resistance in multiple myeloma via ALOX12 signaling. *International Immunopharmacology*, 143(P1), 113320. <https://doi.org/10.1016/j.intimp.2024.113320>
- Elias-Llumbet, A., Sharmin, R., Berg-Sorensen, K., Schirhagl, R., & Mzyk, A. (2024). The Interplay between Mechanoregulation and ROS in Heart Physiology, Disease, and Regeneration. *Advanced Healthcare Materials*, 13(23). <https://doi.org/10.1002/adhm.202400952>
- Feng, Y., Fu, T. X., Zhang, L., Wang, C., & Zhang, D. (2019). Research on differential metabolites in distinction of rice (*Oryza sativa* L.) origin based on GC-MS. *Journal of Chemistry*, 2019. <https://doi.org/10.1155/2019/1614504>
- Fracaschetti, D., Pozzoli, C., Vitalini, S., Tirelli, A., & Iriti, M. (2020). Impact of cooking on bioactive compounds and antioxidant activity of pigmented rice cultivars. *Foods*, 9(8), 1–12. <https://doi.org/10.3390/foods9080967>
- Ghasemzadeh, A., Karbalaii, M. T., Jaafar, H. Z. E., & Rahmat, A. (2018). Phytochemical constituents, antioxidant activity, and antiproliferative properties of black, red, and brown rice bran. *Chemistry Central Journal*, 12(1), 1–13. <https://doi.org/10.1186/s13065-018-0382-9>
- Gong, E. S., Luo, S. J., Li, T., Liu, C. M., Zhang, G. W., Chen, J., Zeng, Z. C., & Liu, R. H. (2017). Phytochemical profiles and antioxidant activity of brown rice varieties. *Food Chemistry*, 227, 432–443. <https://doi.org/10.1016/J.FOODCHEM.2017.01.093>
- Govindarasu, S., & Murugesu, S. (2025). Secondary metabolite based anti-inflammatory, antioxidant activities, drug like properties, molecular docking enzyme inhibitors of selected medicinal plant *Bacopa monnieri* (L)Wettst. *Microbe (Netherlands)*, 8(June), 100478. <https://doi.org/10.1016/j.microb.2025.100478>
- Guo, H., Liu, Z., Li, J., Nie, S., & Pan, W. (2006). Effects of isopropyl palmitate on the skin permeation of drugs. *Biological and Pharmaceutical Bulletin*, 29(11), 2324–2326. <https://doi.org/10.1248/bpb.29.2324>
- Hamilton-Amachree, A., & Uzoekwe, S. A. (2017). GC-MS Analysis of Oil Rich in Polyenoic Fatty Acid Methyl Esters from Leaves of *Justicia Secunda* Vahl Growing Abundantly in The Lowland Rain Forests of The Niger Delta Region of Nigeria. *American Journal of Essential Oils and Natural Products*, 5(4), 1–04.
- Hu, S., Ren, H., Song, Y., Liu, F., Qian, L., Zuo, F., & Meng, L. (2023). Analysis of volatile compounds by GCMS reveals their rice cultivars. *Scientific Reports*, 13(1), 1–10. <https://doi.org/10.1038/s41598-023-34797-2>
- Huang, R., Chen, H., Liang, J., Li, Y., Yang, J., Luo, C., Tang, Y., Ding, Y., Liu, X., Yuan, Q., Yu, H., Ye, Y., Xu, W., & Xie, X. (2021). Dual role of reactive oxygen species and their application in cancer therapy. *Journal of Cancer*, 12(18), 5543–5561. <https://doi.org/10.7150/jca.54699>
- Huang, Y. P., & Lai, H. M. (2016). Bioactive compounds and antioxidative activity of colored rice bran. *Journal of Food and Drug Analysis*, 24(3), 564–574. <https://doi.org/10.1016/j.jfda.2016.01.004>
- Katisart, T., Karirat, T., Saengha, W., Promjamorn, P., Senakun, C., Kaewsri, W., Deeseenthum, S., Somboonwatthanakul, I., Butkhup, L., Sungsi-In, M., Moongngarm, A., Ma, N. L., & Luang-In, V. (2025). Exploring phytochemical profile, antioxidant and anticancer activities of underexploited edible wild plant *Bauhinia saccocalyx* Pierre leaf extract native to Thailand. *Journal of Agriculture and Food Research*, 23(August), 102279. <https://doi.org/10.1016/j.jafr.2025.102279>
- Khromykh, N. O., Lykholat, Y. V., Didur, O. O., Sklyar, T. V., Davydov, V. R., Lavrentieva, K. V., & Lykholat, T. Y. (2022). Phytochemical profiles, antioxidant and antimicrobial activity

- of *Actinidia polygama* and *A. arguta* fruits and leaves. *Biosystems Diversity*, 30(1), 39–45. <https://doi.org/10.15421/012205>
- Krishnaveni, M., Dhanalakshmi, R., & Nandhini, N. (2014). GC-MS analysis of phytochemicals, fatty acid profile, antimicrobial activity of gossypium seeds. *International Journal of Pharmaceutical Sciences Review and Research*, 27(1), 273–276.
- Kusumawardani, S., & Luangsakul, N. (2024). Assessment of polyphenols in purple and red rice bran: Phenolic profiles, antioxidant activities, and mechanism of inhibition against amylolytic enzymes. *Current Research in Food Science*, 9(July), 100828. <https://doi.org/10.1016/j.crfs.2024.100828>
- Li, X., Li, B., Podio, N. S., Wang, X. Y., Jiang, S., Xu, S., Qiu, X., Zeng, Z., Gong, W., Wang, S., & Gong, E. S. (2024). Untargeted metabolomics profiling of purple rice phenolics and their antioxidant activities. *Lwt*, 214(September). <https://doi.org/10.1016/j.lwt.2024.117127>
- Mohebbifar, M. R. (2021). Study of the effect of temperature on thermophysical properties of ethyl myristate by dual-beam thermal lens technique. *Optik*, 247(September), 168000. <https://doi.org/10.1016/j.ijleo.2021.168000>
- Ohiri, R. C., Amadi, B. A., & Onwuzuruike, D. T. (2023). Variations in Confinement of Bioactive Components in Different Sections of *Spondias mombin* Tree. *Journal of Applied Sciences and Environmental Management*, 27(3), 483–493. <https://doi.org/10.4314/jasem.v27i3.11>
- Onoabedje, U. S., Ezugwu, C. O., & Onoabedje, E. A. (2025). Antimicrobial properties of 9, 12-octadecadienoic acid isolated from leaf extracts of *Acalypha fimbriata* (Euphorbiaceae). *Communication in Physical Sciences*, 12(2), 1061–1069. <https://doi.org/10.4314/cps.v12i2.27>
- Pant, P., Pandey, S., & Dall'Acqua, S. (2021). The Influence of Environmental Conditions on Secondary Metabolites in Medicinal Plants: A Literature Review. *Chemistry and Biodiversity*, 18(11). <https://doi.org/10.1002/cbdv.202100345>
- Phanthurat, N., & Thatsanasuwan, N. (2023). A comparative study regrading traditional cooking processes in Northern Thailand influence phytochemical content, antioxidant capacity and inhibition of key enzyme activity in glutinous rice. *Journal of Agriculture and Food Research*, 14(September), 100820. <https://doi.org/10.1016/j.jafr.2023.100820>
- Phuong, L. H., Van Thanh, N., & Binh, L. N. (2025). Volatile compound profile of rice bran extract: Gas chromatography-mass spectrometry optimization and the impact of processing and storage. *Journal of Agriculture and Food Research*, 19(December 2024). <https://doi.org/10.1016/j.jafr.2025.101730>
- Pizzino, G., Irrera, N., Cucinotta, M., Pallio, G., Mannino, F., Arcoraci, V., Squadrito, F., Altavilla, D., & Bitto, A. (2017). Oxidative Stress: Harms and Benefits for Human Health. *Oxidative Medicine and Cellular Longevity*, 2017. <https://doi.org/10.1155/2017/8416763>
- Pranatami, D. A., & Akmalia, H. A. (2025). Potential of baroma rice as anti-cancer food candidate via cell cycle arrest and apoptosis. *Jurnal Teknosains*, 14(2), 126. <https://doi.org/10.22146/teknosains.101243>
- Qin, L., Zhu, W., Yang, L., Zheng, M., & Liu, G. (2025). Persistent free radicals in the environment. *Journal of Hazardous Materials*, 493(March), 310024. <https://doi.org/10.1016/j.jhazmat.2025.138332>
- Saeed, N. M., El-Demerdash, E., Abdel-Rahman, H. M., Algandaby, M. M., Al-Abbasi, F. A., & Abdel-Naim, A. B. (2012). Anti-inflammatory activity of methyl palmitate and ethyl palmitate in different experimental rat models. *Toxicology and Applied Pharmacology*, 264(1), 84–93. <https://doi.org/10.1016/j.taap.2012.07.020>
- Sagna, A., Nair, R. V. R., Hulyalkar, N., Rajasekharan, S., Nair, V. T. G., Sivakumar, K. C., Suja, S. R., Baby, S., & Sreekumar, E. (2023). Ethyl palmitate, an anti-chikungunya virus principle from *Sauropus androgynus*, a medicinal plant used to alleviate fever in ethnomedicine. *Journal of Ethnopharmacology*, 309(January), 116366. <https://doi.org/10.1016/j.jep.2023.116366>
- Shao, Y., Xu, F., Sun, X., Bao, J., & Beta, T. (2014). Identification and quantification of phenolic acids and anthocyanins as antioxidants in bran, embryo and endosperm of white, red and black rice kernels (*Oryza sativa* L.). *Journal of Cereal Science*, 59(2), 211–218. <https://doi.org/10.1016/j.jcs.2014.01.004>
- Singh, N., Mansoori, A., Jiwani, G., Solanke, A. U., Thakur, T. K., Kumar, R., Chaurasiya, M., & Kumar, A. (2021). Antioxidant and antimicrobial study of *Schefflera vinosa* leaves crude extracts against rice pathogens. *Arabian Journal of Chemistry*, 14(7), 103243. <https://doi.org/10.1016/j.arabjc.2021.103243>
- Sokeng, S. D., Talla, E., Sakava, P., Fokam Tagne, M. A., Henoumont, C., Sophie, L., Mbafor, J. T., & Tchuenguem Fohouo, F. N. (2020). Anti-Inflammatory and Analgesic Effect of Arachic Acid Ethyl Ester Isolated from Propolis. *BioMed Research International*, 2020. <https://doi.org/10.1155/2020/8797284>
- Subedi, L., Timalana, S., Duwadi, P., Thapa, R., Paudel, A., & Parajuli, K. (2014). Antioxidant activity and phenol and flavonoid contents of eight medicinal plants from Western Nepal. *Journal of Traditional Chinese Medicine*, 34(5), 584–590. [https://doi.org/10.1016/s0254-6272\(15\)30067-4](https://doi.org/10.1016/s0254-6272(15)30067-4)
- Sulochana, S., Meyyappan, R. M., & Singaravadivel, K. (2016). Phytochemical Screening and Gc-MS Analysis of Garudan Samba Traditional Rice Variety. *International Journal of Environment and Agricultural Research*, 4, 44–47.
- Suryanti, V., Riyatun, Suharyana, Sutarno, & Saputra, O. A. (2020). Antioxidant activity and compound constituents of gamma-irradiated black rice (*Oryza sativa* L.) var. cempo ireng indigenous of Indonesia. *Biodiversitas*, 21(9), 4205–4212. <https://doi.org/10.13057/biodiv/d210935>
- Tang, Y., Cai, W., & Xu, B. (2016). From rice bag to table: Fate of phenolic chemical compositions and antioxidant activities in waxy and non-waxy black rice during home cooking. *Food Chemistry*, 191, 81–90. <https://doi.org/10.1016/j.foodchem.2015.02.001>
- Villanueva-Bermejo, D., Martín Hernández, D., Hurtado-Ribeira, R., Hernández, E. J., Fornari, T., Martín, D., & Vázquez, L. (2025). Supercritical concentration of ethyl laurate from coconut oil. *Lwt*, 225(July 2024). <https://doi.org/10.1016/j.lwt.2025.117928>
- Volpe, C. M. O., Villar-Delfino, P. H., Dos Anjos, P. M. F., & Nogueira-Machado, J. A. (2018). Cellular death, reactive oxygen species (ROS) and diabetic complications review-Article. *Cell Death and Disease*, 9(2). <https://doi.org/10.1038/s41419-017-0135-z>
- Windiyani, H., & Rusdianto, S. W. (2020). Keragaman Varietas Unggul Baru Padi Fungsional Mendukung Ketahanan Pangan dalam Menghadapi Pandemi COVID-19. *Seminar Nasional Lahan* ..., 978–979. <http://www.conference.unsri.ac.id/index.php/lahansuboptimal/article/view/1964>
- Wiraswati, H. L., Fauziah, N., Pradini, G. W., Kurnia, D., Kodir, R. A., Berbudi, A., Arimdayu, A. R., Laelalugina, A., Supandi,

- & Ma'ruf, I. F. (2023). Breynia cernua: Chemical Profiling of Volatile Compounds in the Stem Extract and Its Antioxidant, Antibacterial, Antiplasmodial and Anticancer Activity In Vitro and In Silico. *Metabolites*, 13(2). <https://doi.org/10.3390/metabo13020281>
- Woo, K. S., Kim, H. J., Lee, J. H., Ko, J. Y., Lee, B. W., & Lee, B. K. (2018). Cooking characteristics and antioxidant activity of rice-barley mix at different cooking method and mixing ratio. *Preventive Nutrition and Food Science*, 23(1), 52–59. <https://doi.org/10.3746/pnf.2018.23.1.52>
- Wu, Z., Li, L., Li, N., Zhang, T., Pu, Y., Zhang, X., Zhang, Y., & Wang, B. (2017). Optimization of ultrasonic-assisted extraction of fatty acids in seeds of Brucea javanica (L.) Merr. from different sources and simultaneous analysis using high-performance liquid chromatography with charged aerosol detection. *Molecules*, 22(6), 1–15. <https://doi.org/10.3390/molecules22060931>
- Yusoff, I. M., Mat Taher, Z., Rahmat, Z., & Chua, L. S. (2022). A review of ultrasound-assisted extraction for plant bioactive compounds: Phenolics, flavonoids, thymols, saponins and proteins. *Food Research International*, 157(February), 111268. <https://doi.org/10.1016/j.foodres.2022.111268>